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Fermentation Technology: Use of Various Cultures for Deracemisation of Secondary Alcohols

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Abstract: Enantioselective synthesis of secondary alcohols plays an important role in pharmaceuticals, pheromones, flavors and fragrances, etc. Biocatalysts has unique characteristics when compared with chemical (homogeneous and heterogeneous) catalysts. The present work provides a list of various cultures used for deracemisation/stereoselective synthesis of various natural and synthetic secondary alcohols.

Keywords: Biocatalyst, Enantioselective synthesis, Stereoselective synthesis, Deracemisation, Fermentation technology.

1. Introduction

Development of efficient methods for the synthesis of enantiomerically pure chiral compounds is of tremendous importance. It has been reported that in the year 2004, nine out of ten top drugs contained enantiomerically pure compounds [1]. The requirement of one of the stereospecific products is because it interacts with biological molecules that are highly specific in action. The requirement of enantiomerically pure chiral compounds is not only in the field of pharmaceutics, but also is of great importance in the field of flavor and aroma chemicals, agriculture and specialty materials [2].

Over the past decades, biocatalysis have been recognized as a highly valuable tool for organic chemists and microbiologists to prepare enantiomerically pure molecules. Among the various strategies for the synthesis of chiral molecules, use of chiral catalysts is an important and efficient technique and in enzyme-catalyzed enantioselective synthesis, these biological catalysts are the recently used ones. In addition, biocatalysis is relatively a green technology, as enzyme-catalyzed reactions can be carried out in aqueous medium at ambient temperature and neutral pH under atmospheric pressure and extreme conditions are

also not required thus energy is saved. Moreover, the by-products are also not formed and the reaction is proceeding at an extremely high rate, maybe up to the order of 10^{20} [3]. It has proven to be a useful technology for the chemical industry for the reactions not easily feasible in organic chemistry. These days, highly stereo-, regioand chemoselective biotransformations result in economically attractive, environmentally acceptable and simplified Enzymes manufacturing processes. have been efficiently used for resolution of racemic mixtures, although might not have competed well with chemical methods in synthesis. Phenylethylamine and ibuprofen are commonly cited compounds prepared using enzyme-based chiral processes. The use of pure enzymes requires additional steps that may further complicate the situation, therefore; there is an emerging trend for the use of microbes as a whole for the synthesis of enantiomerically pure compounds. Though, the story of microbial biotransformation has been associated with mankind since long and dates back to around 2000 years BC for the production of vinegar. It is the oldest, best-known examples of oxidations carried out by microbes. Pasteur in 1958 was the first to demonstrate the microbial resolution of tartaric acid. Ammonium salt of tartaric acid was resolved with the

help of mold *Penicillium glaucum*. Pasteur's investigation for the conversion of alcohol into vinegar, also confirmed by Brown was also reported back in the nineteenth century, where the causative organism was named *Bacterium xylinum*.

2. Biocatalyst

Biocatalysts differ from chemical catalysts in many positive ways, viz. selectivity (strict recognition of the substrate with very high enantio-, regio- and chemoselectivities); safety (since reaction conditions are mild, the solvent is water, while ethanol and glucose, etc. are used as hydrogen sources instead of explosive hydrogen gas); natural and environmentally usable (biocatalysts include: microorganisms, plants, animals or their isolated enzymes so can be easily decomposed in the environment after use). Commercially available biocatalysts are easily available or can be cultured easily, including baker's yeast and the alcohol dehydrogenases from baker's yeast, Thermoanaerobium brockii (TBADH), horse liver and the hydroxysteroid dehydrogenase from Pseudomonas testosterone and Bacillus sphaericus.

Biocatalysts can perform a reduction in nonaqueous solvents like ionic liquids, organic solvents like benzene, cyclodextrin and hydrophobic polymers as well. It can be used to control the enantioselectivities as well as to improve the system's environmental friendliness. For example, the alcohol dehydrogenase from *G. candidum* was used in supercritical CO₂ around 100 atm and 35°C and found to catalyze the reduction of fluoroacetophenones. In some cases, there could be competing enzymes with different enantioselectivities, in such condition, an inhibitor of the unnecessary enzyme(s) like allyl bromide, ethyl chloroacetate, allyl alcohol, methyl vinyl ketone, sulfur compounds, Mg²⁺ and Ca²⁺ etc. could be used.

3. Enantioselective Reductions

Enantiomerically pure secondary alcohols are important intermediates for the synthesis of pharmaceuticals, agrochemicals, pheromones, flavors and fragrances, liquid crystals and chiral auxiliaries in asymmetric synthesis [4]. Enzymatic reductions also go via prologs rule [5], but there are incidences where anti pre logs product is formed [6]. The present study is a compendium of work carried out by combining sequential oxidation reductions by the use of microbes to access a range of secondary alcohols from racemates [Table 1].

Microorganism / Cultures	Reactions catalysed	References
Bacillus stearothermophilus, Yarrowia lipolytica	$R \longrightarrow R \longrightarrow OH$ $R = O \longrightarrow OH$ $R = O \longrightarrow OH$ $R = O \longrightarrow OH$ $R \longrightarrow OH$ $O \longrightarrow OH$ OH $O \longrightarrow OH$ OH $O \longrightarrow OH$ OH OH OH OH OH OH OH	[7] & [8]
Alcaligenes bronchisepticus, Streptococcus faecalis	Ph $COOH$ Ph $COOH$ OH $COOH$	[9]
Pseudomonas polycolor, Micrococcus freudenreichii	Ph COOH Ph COOH	[10]
Byssochlamys fulva	$\begin{array}{ccc} & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\$	l [11]
Rhodococcus erythropolis		[12]
N. asteroides, Candida parapsilosis		[13]
Catharanthus roseus	$ \begin{array}{c} & & & & \\ & & & & \\ & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & $	[14]
Candida parapsilosis	ОН ОН ОН	[15]

Table 1.

Candida boidinii	$\bigvee_{i=1}^{OH} \longleftarrow_{i=1}^{OH} \xrightarrow{OH} \longrightarrow_{i=1}^{OH} \xrightarrow{OH} \xrightarrow{OH}$	[16]
Corynespora cassiicola	$\bigcirc \bigcirc $	[17] & [18]
Candida and Pichia spps.	1,2-diols	[17] & [10]
Corynespora cassiicola		[13]
Geotrichum candidum	$\begin{array}{c} R \\ O \\ R = Aryl \operatorname{gp} \end{array} \xrightarrow{R} \xrightarrow{OH} $	[19]
Various bacteria, fungi, and mammalian cultures	Diterpenoids, triterpenoids, steroids and synthetic substrates	[20]
Nicotiana tabacum and Catharanthus roseus	Camphorquinones $\rightarrow \alpha$ -ketoalcohols	[21]
Nocardia pseudosporangifera Nocardia fusca	$ \xrightarrow{OH} \longrightarrow \xrightarrow{OH} \xrightarrow$	[22]
Rhizopus arrhizus	Diarylketone → Diarylmethanol	[23]

4. Conclusion

The production of single enantiomer of various intermediates is increasingly important in the pharmaceutical and other industries. Organic synthesis is one approach for the synthesis of single enantiomer, but the process may be more complicated. Use of biocatalysts provides another dimension for the synthesis of chiral compounds. The resulting advantages of two catalysts over chemical catalysis are stereo- and regioselectivity. Moreover, it is also an aspect of green chemistry, another emerging trend in synthetic chemistry to achieve environmentally friendly processes. Different classes of enzymes provide stereospecificity to different types of reactions, may it be the resolution of racemic compounds or in the asymmetric synthesis of enantiomerically rich chiral compounds. Oxidoreductases have been used in the synthesis of chiral alcohols.

Recently, biological catalysts in the form of whole cells have also been shown to be extremely stereoselective in the synthesis of secondary alcohols. The reduction yield high regio- and stereoselectivity that is difficult to be achieved with chemical catalyst. The need is to identify the particular cultures that will be reaction specific.

References

 Gröger, H. (2010). Enzyme-Catalyzed Asymmetric Synthesis. In Catalytic Asymmetric Synthesis, I. Ojima (Ed.). John Wiley & Sons Inc. pp 269-341.

- [2]. Faber, K. (2000). Biotransformations. Springer, New York.
- [3]. Lehninger, A.L., Nelson, D.L. & Cox, M.M. (2008). Lehninger Principles of Biochemistry. 5th ed., W.H. Freeman, New York.
- [4]. Collins, A.N., Sheldrake, G.N. & Crosby, J. (1997). Chirality in Industry. Wiley and Sons, Chichester.
- [5]. Prelog, V. (1964). Specification of the stereospecificity of some oxidoreductases by diamond lattice sections. *Pure Appl. Chem.*, 9: 119–130.
- [6]. Faber, K. (1997). Biotransformations in Organic Chemistry: A Textbook. 3rd Edn., Springer.
- [7]. Fantin, G., Fogagnolo, M., Giovannini, P.P., Medici, A. & Pedrini, P. (1995). Combined microbial oxidation and reduction: a new approach to the highyield synthesis of homochiral unsaturated secondary alcohols from racemates. *Tetrahedron: Asymmetry*, 6(12): 3047–3053. doi: 10.1016/0957-4166(95)00402-5.
- [8]. Fantin, G., Fogagnolo, M., Medici, A., Pedrini, P. & Fontana, S. (2000). Kinetic resolution of racemic secondary alcohols via oxidation with *Yarrowia lipolytica* strains. *Tetrahedron: Asymmetry*, 11(11): 2367–2373. doi: 10.1016/S0957-4166(00)00185-3.
- [9]. Tsuchiya, S., Miyamoto, K. & Ohta, H. (1992). Highly efficient conversion of (±)-mandelic acid to its (*R*)-(-)-enantiomer by combination of enzyme-mediated oxidation and reduction. *Biotechnol. Lett.*, 14(12): 1137–1142. doi: 10.1007/BF01027017.

- [10]. Takahashi, E., Nakamichi, K. & Furui, M. (1995). *R*-(-)-mandelic acid production from racemic mandelic acids using *Pseudomonas polycolor* IFO 3918 and *Micrococcus freudenreichii* FERM-P 13221. *J. Ferment. Bioeng.*, 80(3): 247–250. doi: 10.1016/0922-338X(95)90824-J.
- [11]. Lanzilotta, R.P., Bradley, D.G. & McDonald, K.M. (1974). Microbial Reduction of Ketopantoyl Lactone to Pantoyl Lactone. *Appl. Microbiol.*, 27(1): 130-134.
- [12]. Shimizu, S., Hattori, S., Hata, H. & Yamada, H. (1987). One-Step Microbial Conversion of a Racemic Mixture of Pantoyl Lactone to Optically Active d-(—)-Pantoyl Lactone. *Appl. Environ. Microbiol.*, 53(3): 519-522.
- [13]. Shimizu, S., Hattori, S., Hata, H. & Yamada, H. (1987). Stereoselective enzymatic oxidation and reduction system for the production of d(–)pantoyl lactone from a racemic mixture of pantoyl lactone. *Enzyme Microb. Technol.*, 9(7): 411–416. doi: 10.1016/0141-0229(87)90136-0.
- [14]. Takemoto, M. & Achiwa, K. (1995). The synthesis of optically active pyridyl alcohols from the corresponding racemates by *Catharanthus roseus* cell cultures. *Tetrahedron: Asymmetry*, 6(12): 2925–2928. doi: 10.1016/0957-4166(95)00388-6.
- [15]. Hasegawa, J., Ogura, M., Tsuda, S., Maemoto, S.i., Kutsuki, H. & Ohashi, T. (1990). High-yield Production of Optically Active 1,2-Diols from the Corresponding Racemates by Microbial Stereoinversion. *Agricultural and Biological Chemistry*, 54(7): 1819–1827. doi: 10.1080/00021369.1990.10870226.
- [16]. Matsumura, S., Kawai, Y., Takahashi, Y. & Toshima, K. (1994). Microbial production of (2R, 4R)-2,4-pentanediol by enatioselective reduction of acetylacetone and stereoinversion of 2,4-

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pentanediol. *Biotechnol. Lett.*, 16(5): 485–490. doi: 10.1007/BF01023330.

- [17]. Carnell, A.J., Iacazio, G., Roberts, S.M. & Willetts, A.J. (1994). Preparation of optically active cyclohexanediols and $(+)-\alpha$ -hydroxycycloheptanone by an enzyme catalysed stereoinversion/oxidation process. *Tetrahedron Lett.*, 35(2): 331–334. doi: 10.1016/S0040-4039(00)76545-2.
- [18]. Goswami, A., Mirfakhrae, K.D. & Patel, R.N. (1999). Deracemization of racemic 1,2-diol by biocatalytic stereoinversion. *Tetrahedron: Asymmetry*, 10(21): 4239–4244. doi: 10.1016/S0957-4166(99)00452-8.
- [19]. Nakamura, K., Kitano, K., Matsuda, T. & Ohno, A. (1996). Asymmetric reduction of ketones by the acetone powder of *Geotrichum candidum*. *Tetrahedron Lett.*, 37(10): 1629–1632. doi: 10.1016/0040-4039(96)00073-1.
- [20]. Asakawa, Y. & Noma, Y. (2010). Biotransformation of Di- and Triterpenoids, Steroids, and Miscellaneous Synthetic Substrates. Comprehensive Natural Products II: Chemistry and Biology, 3: 893–965. doi: 10.1016/B978-008045382-8.00073-3.
- [21]. Chai, W., Hamada, H., Suhara, J. & Akira Horiuchi, C. (2001). Biotransformation of (+)and (-)-camphorquinones by plant cultured cells. *Phytochemistry*, 57(5): 669–673. doi: 10.1016/S0031-9422(01)00133-9.
- [22]. Ogawa, J., Xie, S.-X. & Shimizu, S. (1999). Stereoinversion of optically active 3-pentyn-2-ol by *Nocardia* species. *Biotechnol. Lett.*, 21(4): 331–335. doi: 10.1023/A:1005484731976.
- [23]. Salokhe, P.R., Rashinkar, G.S. & Salunkhe, R.S. (2010). Enantioselective synthesis of diarylmethanols using microbial transformation. *Indian Journal of Chemistry*, 49: 199-202.